Diffuse Midline Gliomas with Histone H3-K27M Mutation: A Series of 47 Cases Assessing the Spectrum of Morphologic Variation and Associated Genetic Alterations

David A. Solomon¹, Matthew D. Wood¹, Tarik Tihan¹, Andrew W. Bollen¹, Nalin Gupta²,³, Joanna J. Phillips¹,², and Arie Perry¹,²

1. Division of Neuropathology, Department of Pathology, University of California, San Francisco
2. Department of Neurological Surgery, University of California, San Francisco
3. Department of Pediatrics, University of California, San Francisco

To whom correspondence should be addressed: Dr. Arie Perry, Division of Neuropathology, University of California, San Francisco, 505 Parnassus Ave, Room M-551, San Francisco, CA 94143, United States, Phone: 415-476-5236, Fax: 415-476-7963, Email: arie.perry@ucsf.edu

Keywords: histone H3.1, histone H3.3, H3F3A, HIST1H3B, K27M mutation, astrocytoma, glioblastoma, diffuse midline glioma, diffuse intrinsic pontine glioma, DIPG
ABSTRACT

Somatic mutations of the $H3F3A$ and $HIST1H3B$ genes encoding the histone H3 variants, H3.3 and H3.1, were recently identified in high-grade gliomas arising in the thalamus, pons, and spinal cord of children and young adults. However, the complete range of patients and locations in which these tumors arise, as well as the morphologic spectrum and associated genetic alterations remain undefined. Here we describe a series of 47 diffuse midline gliomas with histone H3-K27M mutation. The 25 male and 22 female patients ranged in age from 2 to 65 years (median=14). Tumors were centered not only in the pons (17), thalamus (15), and spinal cord (9), but also in the third ventricle (3), hypothalamus (1), pineal region (1), and cerebellum (1). Patients with pontine tumors were younger (median=7 years) than those with thalamic (median=24 years) or spinal (median=25 years) tumors. A wide morphologic spectrum was encountered including gliomas with giant cells, epithelioid and rhabdoid cells, primitive neuroectodermal tumor (PNET)-like foci, neuropil-like islands, pilomyxoid features, ependymal-like areas, sarcomatous transformation, ganglionic differentiation, and pleomorphic xanthoastrocytoma (PXA)-like areas. In this series, histone H3-K27M mutation was mutually exclusive with $IDH1$ mutation and $EGFR$ amplification, rarely co-occurred with $BRAF-V600E$ mutation, and was commonly associated with p53 overexpression, ATRX loss (except in pontine gliomas), and monosomy 10.
INTRODUCTION

Recent genomic analysis has demonstrated that specific genetic alterations drive distinct subsets of glial neoplasms of the central nervous system, dependent not only on tumor type but also site of origin and patient age. For example, two independent studies in 2012 reported the identification of somatic mutation of the H3F3A and HIST1H3B genes, which encode the histone H3 variants H3.3 and H3.1, in pediatric diffuse intrinsic pontine gliomas (DIPG) and non-brainstem glioblastomas (25, 29). A recurrent lysine to methionine substitution at codon 27 (K27M) was present in one of these two histone H3 variants in the majority of DIPGs and thalamic glioblastomas, whereas a recurrent glycine to arginine or valine substitution at codon 34 (G34R/V) was present in a subset of those pediatric non-midline glioblastomas arising the cerebral hemispheres. Subsequent analyses have now demonstrated that histone H3-K27M mutations are present in the majority of high grade infiltrative astrocytomas arising within midline structures (thalamus, pons, and spinal cord) of both pediatric and young adult patients (1, 5, 9, 11, 12, 19, 20, 24, 26). Studies have shown that these diffuse midline gliomas with histone H3-K27M mutation are associated with aggressive clinical behavior and poor prognosis (19, 20, 26), including those tumors which demonstrate only low grade histologic features on biopsy (5). However, more recent studies have shown that thalamic gliomas in adult patients with histone H3-K27M mutation might not be associated with worse prognosis than corresponding histone H3 wild-type thalamic gliomas (1, 9), suggesting heterogeneity amongst this molecular subgroup of diffuse midline gliomas.

Investigation has shown that this K27M mutation alters an important site of post-translational modification in the histone H3 variants and leads to altered DNA methylation and gene expression profiles thought to drive gliomagenesis (3, 7, 15, 21, 26). Thus, there are ongoing efforts to study the efficacy of therapeutics targeting histone modifying enzymes for these midline gliomas with histone H3 mutations. A recent preclinical study demonstrated remarkable efficacy with a novel small molecule inhibitor of the histone demethylase JMJD3 both in vitro and in vivo orthotopic xenograft models (14). A second group
recently demonstrated therapeutic efficacy with the histone deacetylase inhibitor panobinostat both in vitro and in vivo orthotopic xenograft models (13). It is thus important to identify those gliomas with histone H3 mutations, given the prognostic significance and potential targeted therapy that will soon be in clinical trials for these patients.

Genomic analysis of diffuse midline gliomas with histone H3-K27M mutations (K27M+) has revealed a number of cooperating genetic alterations. In particular, these tumors often also have TP53 and ATRX mutations but do not have IDH1 mutation (19, 25, 26). More recently, a subset of K27M+ DIPGs were found to also harbor missense mutations in the ACVR1 gene, encoding the activin A receptor type-1 transmembrane protein, that lead to activation of the BMP-TGFβ signaling pathway (6, 10, 27, 30). Other alterations occasionally found in K27M+ DIPGs include PIK3CA mutation, PDGFRA mutation or amplification, PPM1D mutation, and amplification of cell cycle genes including CCND1, CDK4, and CDK6 (30, 31). The cooperating genetic alterations in non-brainstem gliomas with histone H3-K27M mutation are less well defined, particularly in spinal cord and thalamic gliomas in adult patients. Notably, rare examples of midline glial neoplasms have been identified in pediatric patients that harbor both histone H3-K27M mutation and BRAF-V600E mutation (23). Accurate pathologic diagnosis in these cases was reportedly difficult, and two patients had long term survival of three or more years after diagnosis.

Additionally, the morphologic spectrum of K27M+ diffuse midline gliomas has only been reported in a limited fashion. In one assessment of 6 pediatric epithelioid glioblastomas, one tumor centered in the thalamus harbored a K27M mutation in H3F3A, suggesting that epithelioid features can occasionally be seen (4). Additionally, a recent case report described a tumor with histologic features resembling pilocytic astrocytoma arising in the cervical spinal cord of a 7 year old girl that harbored histone H3-K27M mutation but lacked the characteristic BRAF-KIAA1549 gene fusion (16). This patient had disease-free survival for 10 years following resection without additional therapy but subsequently had disease...
recurrence with malignant transformation causing her eventual death 12 months later. Such cases suggest that this molecular subgroup of diffuse midline gliomas is not morphologically uniform.

In this study, we report clinicopathologic assessment of 73 infiltrative gliomas and 15 pilocytic astrocytomas centered in midline structures in both pediatric and adult patients, including immunohistochemistry with a recently developed mutant-specific antibody that accurately detects the K27M mutation of both histone H3.3 and histone H3.1 (2, 28). Our findings highlight the range of midline locations and patients in which gliomas with histone H3-K27M mutation arise, as well as demonstrate the diverse histopathologic spectrum that can be observed in this molecular subgroup of diffuse midline gliomas.
MATERIAL AND METHODS

Tumor samples and histologic review

This study was performed in accordance with guidelines set forth by the Institutional Review Board of the University of California, San Francisco (UCSF). For all tumors, formalin-fixed paraffin-embedded tumor tissue was prospectively examined from both UCSF in-house and consult cases over a one year period. Hematoxylin and eosin (H&E) stained sections were reviewed for diagnostic assessment and determination of morphologic variation. Amongst the 47 K27M+ diffuse midline gliomas, 17 of the pediatric cases and 10 of the adult cases were resected in-house, whereas 10 of the pediatric cases and 10 of the adult cases were resected at outside institutions and reviewed at UCSF in consultation.

Immunohistochemistry and fluorescence in situ hybridization (FISH)

Immunohistochemistry was performed on formalin-fixed, paraffin embedded tissue sections at the UCSF Immunohistochemistry Laboratory and the UCSF Neuropathology BTRC Biomarkers Laboratory. Primary antibodies used were as follows: histone H3-K27M mutant protein (ABE419, EMD Millipore, Billerica, MA, 1:500 dilution), IDH1-R132H mutant protein (clone H09, DiaNova, Germany, 1:500 dilution), ATRX (HPA001906, Sigma, St Louis, MO, 1:100 dilution), p53 (clone DO-7, Dako, Glostrup, Denmark, 1:00 dilution), BRAF-V600E mutant protein (clone VE1, Ventana, Tucson, AZ). All staining was performed in Ventana or Leica Bond automated staining processors. Specifically, the histone H3-K27M immunostaining was run in a Ventana Benchmark XT autostainer using CC1 antigen retrieval buffer for 30 min at 95°C, incubation with 1:500 dilution of primary antibody for 32 min, and Ventana ultraView Universal DAB detection. Dual-color FISH for EGFR and CEP7 or PTEN and CEP10 was performed on 5-micron thick FFPE whole sections as previously described (17).
RESULTS

Immunohistochemistry for histone H3-K27M mutant protein was performed on a total of 73 infiltrative gliomas arising in midline locations, 21 infiltrative gliomas arising in the cerebral hemispheres, and 15 pilocytic astrocytomas arising in midline locations (Table 1). 47 of the 73 infiltrative gliomas (64%) arising in midline locations were positive for histone H3-K27M mutant protein, while none of the 21 infiltrative gliomas arising in the cerebral hemispheres or 15 pilocytic astrocytomas were positive.

Among the midline infiltrative gliomas, histone H3-K27M mutant protein was detected in 17 of 18 (94%) pontine, 15 of 23 (65%) thalamic, 9 of 17 (53%) spinal, 3 of 3 (100%) third ventricular, 1 of 4 (25%) hypothalamic, 1 of 4 (25%) cerebellar, 1 of 2 (50%) pineal, 0 of 1 (0%) midbrain (0%), and 0 of 1 (0%) medulla-centered gliomas.

Clinical features of diffuse midline gliomas with histone H3-K27M mutation

Among the 47 patients with diffuse midline gliomas positive for histone H3-K27M mutation, 25 were males and 22 were females (Table 2). Patient age at time of diagnosis ranged from 2 to 65 years old with a median age of 14 years. Patients with pontine gliomas were younger (median=7 years) than those with thalamic (median=24 years) or spinal (median=25 years) gliomas (Figure 1).

Thirty four of these 47 patients underwent biopsy only, whereas 13 underwent resection (Table 2). Initial histologic diagnosis was diffuse astrocytoma (WHO grade II) in 10 cases, anaplastic astrocytoma (WHO grade III) in 14 cases, and glioblastoma (WHO grade IV) in 23 cases. During the short period of clinical follow-up, five of these patients underwent a subsequent biopsy or resection demonstrating higher grade gliomas. These included two pontine gliomas originally classified as grade III that were upgraded to glioblastoma, a spinal glioma and a third ventricle glioma originally classified as grade II that were upgraded to glioblastoma, and a thalamic glioma originally classified as grade II that was upgraded to anaplastic astrocytoma.
Given the prospective nature of this study without significant length of clinical follow-up, the utility of assessing outcome measures including recurrence-free survival and overall survival in this patient cohort is limited at present. A follow-up study will assess these clinical parameters to evaluate for differences in clinical outcome based on histone H3-K27M mutation status, tumor site, patient age, associated molecular alterations, and other variables in this patient cohort.

**Histologic features of diffuse midline gliomas with histone H3-K27M mutation**

In assessing the morphologic features of these 47 diffuse midline gliomas with histone H3-K27M mutation, all tumors had at least some classic astrocytic morphology with ovoid to elongate nuclei containing coarse chromatin. None had widespread oligodendroglial morphology with uniform round nuclei and more delicate chromatin warranting classification as oligodendroglioma. In each case, histone H3-K27M mutant protein staining was diffusely positive throughout all tumor nuclei, suggesting that histone H3 mutation is an early or initiating event in these diffuse midline gliomas. Additionally, histone H3-K27M positivity was limited to the neoplastic cells in all cases, with no nuclear staining seen in endothelial cells, inflammatory cells, or entrapped non-neoplastic neurons and glial cells. As a potential diagnostic pitfall, nonspecific cytoplasmic staining was often seen in admixed macrophages and microglial cells in tumors lacking histone H3-K27M mutant protein (Figure 2), although this pattern was easy to distinguish from the nuclear staining seen in positive tumor cells. A representative example of a diffuse midline glioma with histone H3-K27M mutation arising in the pons of an 8 year old boy is shown in Figure 3. An autopsy was performed on this patient following palliative radiation and chemotherapy, and histone H3-K27M immunohistochemistry was helpful in assessing the widespread infiltration of the tumor cells throughout the brain and spinal cord. Tumor cells were seen infiltrating from the brainstem into multiple cranial nerves (Fig. 3H-I), a finding that has only been rarely observed in malignant gliomas (22).
While the vast majority of diffuse midline gliomas with histone H3-K27M mutation previously reported have been centered in the thalamus, pons, or spinal cord, we identified 6 malignant gliomas arising in other midline sites including the third ventricle, hypothalamus, pineal region, and cerebellum (Figure 4). All of these tumors displayed typical morphologic features of infiltrative astrocytomas, except for one glioblastoma arising in the third ventricle of a 19 year old woman with tumor cells that contained clear cytoplasm and distinct cell borders (Fig. 4G-I).

Among the 41 diffuse midline gliomas with histone H3-K27M mutation arising in the thalamus, pons, and spinal cord, a wide spectrum of morphologic variation was encountered (Figures 5-7). Two glioblastomas had frequent giant cells, including one arising in the pons of a 7 year old girl and one arising in the thalamus of a 12 year old girl (Fig. 5A-B), both of which showed p53 overexpression in the majority of tumor nuclei. One glioblastoma arising in the cervical spinal cord of a 15 year old boy demonstrated prominent pilomyxoid features with rather monomorphic piloid astrocytes radially arranged vessels in a prominent myxoid stroma, resembling pilomyxoid astrocytoma. Other areas of the tumor demonstrated increased cellularity and pleomorphism with brisk mitotic activity and microvascular proliferation, which together with the histone H3-K27M positivity, warranted classification as glioblastoma, WHO grade IV (Fig. 5C-D). Another notable glioblastoma arising in the thalamus of a 27 year old woman demonstrated epithelioid features and also contained primitive neuroectodermal tumor (PNET)-like foci (Fig. 5E-H).

Another interesting glioma variant we observed to harbor histone H3-K27M mutation was a glioblastoma with neuropil-like islands arising in the thoracic spinal cord of a 25 year old woman (Fig. 6A-C). This tumor contained neuropil-like islands as highlighted by synaptophysin immunohistochemistry circumferentially surrounded by neoplastic astrocytes. Other areas of the tumor contained necrosis and microvascular proliferation, warranting classification as glioblastoma. As opposed to the vast majority of the diffuse gliomas with neuropil-like islands that have been reported to date (18), this tumor lacked IDH1 mutant protein. One gliosarcoma arising in the thalamus of an 11 year old girl highlighted the spectrum
of intratumoral variation that can be observed in diffuse midline gliomas with histone H3-K27M mutation. Portions of this tumor were composed of ependymal-like areas with solid growth, uniform ovoid nuclei, no mitotic activity, abundant dystrophic calcification, perivascular pseudorosettes, and cytoplasmic dot-like immunohistochemical staining for EMA (Fig. 6D-F). These low grade areas were adjacent to more anaplastic appearing areas composed of spindled cells with fascicular growth, brisk mitotic activity, and loss of markers of glial differentiation such as GFAP and OLIG2 suggestive of sarcomatous transformation (Fig. 6G-I). Another morphologic variant was a malignant glioma with epithelioid to rhabdoid features and ganglionic differentiation in the thoracic spinal cord of a 26 year old man. The majority of this tumor was composed of loosely cohesive cells with eccentric nuclei and abundant eosinophilic cytoplasm, many of which contained spherical intracytoplasmic inclusions (Fig. 6J-L). One area of the tumor demonstrated numerous large atypical ganglion-like cells which were also positive for histone H3-K27M mutant protein (Fig. 6M-O). The classification of this tumor remained enigmatic, though we favored that it represents a diffuse midline glioma with epithelioid/rhabdoid features and ganglionic differentiation rather than an anaplastic ganglioglioma with malignant glial component demonstrating features of epithelioid/rhabdoid glioblastoma.

**Molecular features of diffuse midline gliomas with histone H3-K27M mutation**

As part of the clinical workup of these 47 diffuse midline gliomas with histone H3-K27M mutation, a combination of additional molecular testing was variably performed including immunohistochemistry for ATRX, p53, IDH1-R132H mutant protein, and BRAF-V600E mutant protein, as well as fluorescence in situ hybridization (FISH) for EGFR and PTEN genes (Table 2). ATRX immunostaining revealed loss of nuclear expression in 15 of 32 tested cases, including 9 of 12 thalamic gliomas (75%), 4 of 7 spinal gliomas (57%), and 1 of 13 pontine gliomas (8%). Of the 33 cases assessed, 14 demonstrated strong p53 immunoreactivity in the majority of tumor nuclei suggestive of TP53 gene mutation or other mechanism.
of p53 stabilization. Whereas ATRX loss was more commonly seen in the thalamic and spinal gliomas, p53 overexpression was more frequent in the thalamic (8 of 13 cases, 62%) versus pontine (4 of 12 cases, 33%) and spinal gliomas (1 of 5 cases, 20%). No significant correlation between tumor grade and ATRX or p53 status was seen in this cohort. Of the 30 cases tested for IDH1-R132H mutant protein, none were positive. Only one of the 18 cases tested for BRAF-V600E mutant protein was positive, which was the exceptional glioma arising in the thalamus of a 5 year old girl depicted in Figure 7. This case demonstrated areas of both a diffusely infiltrative glioma admixed with other solid areas resembling PXA, including nuclear pleomorphism out of proportion to mitotic activity, dense reticulin network, eosinophilic granular bodies, and CD34 positive stellate cells. Of 15 cases assessed by FISH, none had EGFR amplification while four had loss of chromosome 10, including a pontine glioblastoma in an 11 year old boy, a pontine glioblastoma in a 4 year old boy, a thalamic glioblastoma in a 24 year old woman, and a glioma in the lumbar spinal cord with grade II histologic features at time of initial biopsy but grade IV histologic features on subsequent resection.

Characteristics of diffuse midline gliomas without histone H3-K27M mutation

26 of the 73 diffuse midline gliomas evaluated were negative for histone H3-K27M mutant protein expression. These included 1 of 18 tumors (6%) arising in the pons, 8 of 23 (35%) arising in the thalamus, 8 of 17 (47%) arising in the spinal cord, 3 of 4 (75%) arising in the hypothalamus, 3 of 4 (75%) arising in the cerebellum, 1 of 2 (50%) arising in the pineal region, 1 of 1 (100%) arising in the midbrain, and 1 of 1 (100%) arising in the medulla. Two of these tumors (a glioblastoma in the thalamus of a 40 year old woman and a glioblastoma in the hypothalamus of a 57 year old man) had EGFR amplification and chromosome 10 deletion, similar to those more typically arising in the cerebral hemispheres. However, the molecular underpinnings of the rest of these tumors remains unknown, and none of the 19 tested cases had IDH1 mutation.
Notably, three of these diffuse midline gliomas lacking histone H3-K27M mutation had a history of prior radiation therapy. These included the only K27M negative DIPG in this study that occurred in a 15 year old boy after receiving craniospinal irradiation at 5 years of age following resection of a medulloblastoma from the midline of the posterior fossa. The second patient was a 35 year old woman with glioblastoma centered in the thalamus who had received focused radiation to the pineal region four years prior following resection of a pineal parenchymal tumor. The third patient was a 46 year old woman with a glioblastoma arising in the thoracic spinal cord following prior radiation therapy for Hodgkin’s lymphoma.
DISCUSSION

“Diffuse midline glioma with histone H3-K27M mutation” is a distinct subtype of infiltrative glioma that will be recognized as a new diagnostic entity in the forthcoming 2016 edition of the *World Health Organization Classification of Tumors of the Central Nervous System* that utilizes integrated diagnosis incorporating both morphologic and molecular features. Emerging evidence indicates that these tumors arise in the thalamus, pons, and spinal cord of children and young adults and are associated with poor prognosis. Given the prognostic implications and the new targeted therapies being developed for these gliomas with histone H3 gene mutations, there is increasing importance for recognition of this distinct glioma entity by diagnostic pathologists. We sought to determine the complete range of patients and locations in which these gliomas with histone H3-K27M mutation arise, as well as the morphologic spectrum and associated genetic alterations that can be seen in these tumors.

We identified 47 patients with diffuse midline gliomas harboring histone H3-K27M mutation. While all prior examples have been in pediatric and young adult patients, we found K27M+ gliomas in older adults up to 65 years of age. While nearly all prior examples of diffuse gliomas with histone H3-K27M mutation have been located in the thalamus, pons, or spinal cord, we also found examples arising in the third ventricle, hypothalamus, pineal region, and cerebellum. Furthermore, we note that a wide morphologic spectrum can be encountered, with features overlapping nearly every known histologic variant of infiltrative glioma and even some of the more circumscribed subtypes such as ganglioglioma and PXA.

In this series of 47 diffuse midline gliomas, histone H3-K27M mutation was mutually exclusive with *IDH1*-R132H mutation and *EGFR* amplification, rarely co-occurred with *BRAF-V600E* mutation, and was commonly associated with p53 overexpression, ATRX loss, and monosomy 10. Amongst these K27M+ diffuse midline gliomas, ATRX loss was more common in thalamic and spinal tumors versus pontine tumors, whereas p53 overexpression was most frequent in thalamic gliomas compared to the other sites.
Whether these molecular differences between different anatomic locations reflects distinct glioma subgroups with divergent clinical outcomes remains to be determined, but this finding further substantiates the concept that K27M+ diffuse midline gliomas are a heterogeneous group of tumors, both morphologically and genetically.

We identified no diffuse gliomas arising in the cerebral hemispheres and no midline pilocytic astrocytomas with histone H3-K27M mutation. A subset of the diffuse midline gliomas we assessed were negative for histone H3-K27M mutant protein. While some of these tumors had molecular features similar to those seen in glioblastomas arising in the cerebral hemispheres (e.g. \textit{EGFR} amplification, \textit{PTEN} deletion), the majority lacked known glioma molecular alterations including \textit{EGFR} amplification, \textit{IDH1} mutation, and ATRX loss. These K27M negative diffuse midline gliomas included all of the three radiation-associated gliomas evaluated in this study, providing further support for the concept that radiation-induced gliomas are driven by a set of genetic alterations distinct from sporadic gliomas (8).

In summary, we recommend a liberal use of immunohistochemistry for clinical detection of histone H3-K27M mutation in all diffuse midline gliomas regardless of patient age. Given that diffuse midline gliomas with histone H3-K27M mutation can display a wide spectrum of morphologic variation, this immunostain may also be useful in midline glial and glioneuronal neoplasms with unusual morphologic patterns that would not previously have been considered part of the diffuse midline glioma family of tumors.
ACKNOWLEDGEMENTS

We thank the UCSF Neuropathology BTRC Biomarkers Laboratory for developing and performing the immunohistochemical stain for histone H3-K27M mutant protein. D.A.S. is supported by a Career Development Award from the UCSF Brain Tumor SPORE and an NIH Director’s Early Independence Award (DP5 OD021403).
REFERENCES


FIGURE LEGENDS

Figure 1. Age distribution of the 47 patients with diffuse midline gliomas with histone H3-K27M mutation at time of initial diagnosis plotted by tumor site. Pons, 17 cases; thalamus, 15 cases; spinal cord, 9 cases; other midline sites, 6 cases.

Figure 2. Example of a diffuse midline glioma negative for histone H3-K27M mutation. (A) H&E stained section of a glioblastoma arising in the medulla of a 46 year old man. (B) Immunohistochemistry for histone H3-K27M mutant protein showed non-specific cytoplasmic staining in admixed microglia and macrophages with no staining of tumor nuclei.

Figure 3. Pathologic features of a diffuse midline glioma with histone H3-K27M mutation arising in the pons in an 8 year old boy from the initial biopsy and at autopsy following palliative radiation and chemotherapy. (A) Mid-sagittal T2 FLAIR image at time of presentation demonstrating a non-enhancing, T2-hyperintense mass expanding the pons with probable infiltration into the adjacent cerebellum. (B) H&E stained section from a diagnostic biopsy performed at the time of presentation showing a highly cellular infiltrative astrocytoma with scattered mitotic figures. (C) Immunohistochemical stain for histone H3-K27M mutant protein showing strong nuclear positivity in tumor cells. (D-E) Gross photographs of an axial section through the pons (D) and coronal section through the cerebellum (E) from the autopsy performed 12 months after the initial diagnostic biopsy following palliative radiation and chemotherapy with bevacizumab. The pons and cerebellum are markedly expanded and distorted by the tumor and show multiple foci of radiation-induced necrosis. (F-G) H&E stained section from the left cerebellar cortex and corresponding immunohistochemical stain for histone H3-K27M mutant protein showing massive infiltration by neoplastic astrocytes with prominent subpial accumulation. (H) Immunohistochemical stain
for histone H3-K27M from the midbrain highlights cranial nerve infiltration by the tumor cells. (I) H&E stained cross-section of a cranial nerve showing numerous neoplastic astrocytes within the endoneurium.

**Figure 4.** Examples of diffuse midline gliomas with histone H3-K27M mutation arising in midline sites other than the thalamus, pons, or spinal cord. (A-C) Glioblastoma arising in the cerebellum of a 9 year old boy. (A) Pre-operative axial T1 post-contrast MR image demonstrating a peripherally enhancing mass within the left cerebellar white matter and cerebellar peduncle. H&E stained section showing a highly pleomorphic infiltrating astrocytoma with microvascular proliferation (B), and immunohistochemical stain for histone H3-K27M mutant protein showing strong nuclear positivity in tumor cells (C). (D-F) Anaplastic astrocytoma centered in the pineal region of a 65 year old man. (D) Pre-operative axial post-contrast CT image demonstrating a faintly enhancing mass in the pineal region. H&E stained section showing an infiltrative astrocytoma with scattered mitotic figures (E), and immunohistochemical stain for histone H3-K27M mutant protein showing strong nuclear positivity in tumor cells (F). (G-I) Glioblastoma arising in the third ventricle in a 19 year old woman. (G) Coronal T1 post-contrast MR image taken following stereotactic biopsy showing a peripherally enhancing mass centered in the third ventricle near the foramen of Monro. H&E stained section showing an infiltrative astrocytoma with clear cytoplasm and well developed microvascular proliferation (H), and immunohistochemical stain for histone H3-K27M mutant protein showing strong nuclear positivity in tumor cells.

**Figure 5.** Examples of morphologic variation encountered in diffuse midline gliomas with histone H3-K27M mutation. (A-B) Glioblastoma with frequent giant cells arising in the thalamus of a 12 year old girl. (C-D) Glioblastoma with pilomyxoid features arising in the cervical spinal cord of a 15 year old boy which demonstrates monomorphic piloid astrocytes radially arranged vessels in a prominent myxoid stroma. Other areas of the tumor demonstrated increased cellularity and pleomorphism with brisk mitotic activity.
and microvascular proliferation (not shown).  **(E-H)** Epithelioid glioblastoma with primitive neuroectodermal tumor (PNET)-like foci arising in the thalamus of a 27 year old woman. The majority of the tumor was composed of epithelioid astrocytes with abundant eosinophilic cytoplasm and distinct cell borders (E), with a smaller component of PNET-like foci composed of more primitive appearing cells with scant cytoplasm, nuclear molding, and brisk mitotic activity (G). H&E stained sections are shown in A, C, E, and G, and corresponding immunohistochemical stained sections for histone H3-K27M mutant protein are shown in B, D, F, and H.

**Figure 6.** Additional examples of morphologic variation encountered in diffuse midline gliomas with histone H3-K27M mutation.  **(A-C)** Glioblastoma with neuropil-like islands arising in the thoracic spinal cord of a 25 year old woman. The tumor contained frequent neuropil-like islands, highlighted by synaptophysin immunohistochemistry (B), circumferentially surrounded by neoplastic astrocytes. Other areas of the tumor contained necrosis and microvascular proliferation (not shown).  **(D-I)** Gliosarcoma with ependymal-like areas arising in the thalamus of an 11 year old girl. Areas within the tumor showed solid growth with uniform ovoid nuclei, no mitotic activity, and perivascular arrangement resembling the pseudorosettes seen in ependymomas (D). Immunohistochemical staining for epithelial membrane antigen (EMA) in these areas of the tumor demonstrated dot-like cytoplasmic positivity (E). Other areas showed a more anaplastic sarcoma-like component with fascicles of spindled cells and brisk mitotic activity (G) that demonstrated increased intercellular reticulin deposition (not shown) and loss of markers of glial differentiation, including OLIG2 (H). Strong nuclear staining for histone H3-K27M mutant protein was seen diffusely throughout the tumor including both the ependymal-like (F) and sarcomatous components (I).  **(J-O)** Malignant glioma with epithelioid/rhabdoid features and ganglionic differentiation arising in the thoracic spinal cord of a 26 year old man. The majority of the tumor was composed of loosely cohesive cells with eccentric nuclei and abundant eosinophilic cytoplasm, many of which contain spherical
intracytoplasmic inclusions (J). Immunohistochemistry for MAP2 showed cytoplasmic staining in the majority of cells with absence of staining in these paranuclear inclusions (K). One area of the tumor demonstrated numerous large atypical ganglion-like cells (M), which demonstrated strong cytoplasmic positivity on MAP2 immunostaining (N), as well as immunoreactivity for neurofilament and synaptophysin (not shown). Strong nuclear staining for histone H3-K27M mutant protein was seen diffusely throughout the tumor including both the epithelioid/rhabdoid cells (L) and the ganglion-like tumor cells (O).

**Figure 7.** Diffuse midline glioma with focal pleomorphic xanthoastrocytoma (PXA)-like features and both histone H3-K27M and **BRAF-V600E** mutation arising in the thalamus of a 5 year old girl. **(A)** Pre-operative coronal T1 post-contrast MR image demonstrating a complex, enhancing multinodular mass centered in the left thalamus with infiltration into the surrounding brain parenchyma. **(B,D)** A large component of the tumor demonstrated histologic features of pleomorphic xanthoastrocytoma (PXA) with pleomorphic astrocytes, fascicular to storiform architecture, solid growth pattern, numerous eosinophilic granular bodies, and occasional xanthomatous cells. These PXA-like areas were in apposition to a diffusely infiltrative astrocytoma with scattered mitotic figures and infarct-like necrosis not associated with palisading or microvascular proliferation (C). The diffusely infiltrative component of the tumor lacked significant reticulin deposition (E) and had CD34 immunostaining limited to endothelial cells (G), while the PXA-like areas had a dense intercellular reticulin network (F) and interspersed stellate cells with CD34 staining (H). Staining for both **BRAF-V600E** and histone H3-K27M mutant proteins was seen diffusely throughout the tumor including both the infiltrative component (I,K) and the PXA-like component (J,L).
Table 1. IHC results for histone H3-K27M mutant protein.

<table>
<thead>
<tr>
<th>Infiltrative gliomas</th>
<th>K27M+</th>
<th>K27M-</th>
<th>Total</th>
<th>% K27M+</th>
</tr>
</thead>
<tbody>
<tr>
<td>Pons</td>
<td>17</td>
<td>1</td>
<td>18</td>
<td>94%</td>
</tr>
<tr>
<td>Thalamus</td>
<td>15</td>
<td>8</td>
<td>23</td>
<td>65%</td>
</tr>
<tr>
<td>Spinal cord</td>
<td>9</td>
<td>8</td>
<td>17</td>
<td>53%</td>
</tr>
<tr>
<td>Third ventricle</td>
<td>3</td>
<td>0</td>
<td>3</td>
<td>100%</td>
</tr>
<tr>
<td>Hypothalamus</td>
<td>1</td>
<td>3</td>
<td>4</td>
<td>25%</td>
</tr>
<tr>
<td>Cerebellum</td>
<td>1</td>
<td>3</td>
<td>4</td>
<td>25%</td>
</tr>
<tr>
<td>Pineal region</td>
<td>1</td>
<td>1</td>
<td>2</td>
<td>50%</td>
</tr>
<tr>
<td>Cerebral hemispheres</td>
<td>0</td>
<td>21</td>
<td>21</td>
<td>0%</td>
</tr>
<tr>
<td>Midbrain</td>
<td>0</td>
<td>1</td>
<td>1</td>
<td>0%</td>
</tr>
<tr>
<td>Medulla</td>
<td>0</td>
<td>1</td>
<td>1</td>
<td>0%</td>
</tr>
<tr>
<td>Total for midline sites</td>
<td>47</td>
<td>26</td>
<td>73</td>
<td>64%</td>
</tr>
<tr>
<td>Total for all sites</td>
<td>47</td>
<td>47</td>
<td>94</td>
<td>50%</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Pilocytic astrocytomas</th>
<th></th>
<th></th>
<th></th>
<th></th>
</tr>
</thead>
<tbody>
<tr>
<td>Spinal cord</td>
<td>0</td>
<td>5</td>
<td>5</td>
<td>0%</td>
</tr>
<tr>
<td>Cerebellum</td>
<td>0</td>
<td>4</td>
<td>4</td>
<td>0%</td>
</tr>
<tr>
<td>Tectum</td>
<td>0</td>
<td>3</td>
<td>3</td>
<td>0%</td>
</tr>
<tr>
<td>Third ventricle</td>
<td>0</td>
<td>1</td>
<td>1</td>
<td>0%</td>
</tr>
<tr>
<td>Hypothalamus</td>
<td>0</td>
<td>1</td>
<td>1</td>
<td>0%</td>
</tr>
<tr>
<td>Medulla</td>
<td>0</td>
<td>1</td>
<td>1</td>
<td>0%</td>
</tr>
<tr>
<td>Total for all sites</td>
<td>0</td>
<td>15</td>
<td>15</td>
<td>0%</td>
</tr>
</tbody>
</table>
Table 2. Clinical and molecular features of diffuse midline gliomas with histone H3-K27M mutation.

<table>
<thead>
<tr>
<th>Tumor site</th>
<th>Cases</th>
<th>Sex (M:F)</th>
<th>Median</th>
<th>Range</th>
<th>Biopsy</th>
<th>Resection</th>
<th>WHO grade</th>
<th>Immunohistochemistry</th>
<th>FISH</th>
</tr>
</thead>
<tbody>
<tr>
<td>Pons</td>
<td>17</td>
<td>8:9</td>
<td>7</td>
<td>4-24</td>
<td>16</td>
<td>1</td>
<td>5</td>
<td>8 4 0/10 1/13 4/12 0/7</td>
<td>0/4 2/4</td>
</tr>
<tr>
<td>Thalamus</td>
<td>15</td>
<td>7:8</td>
<td>24</td>
<td>2-60</td>
<td>11</td>
<td>4</td>
<td>1 4 10</td>
<td>0/11 9/12 8/13 1/4</td>
<td>0/6 1/6</td>
</tr>
<tr>
<td>Spinal cord</td>
<td>9</td>
<td>5:4</td>
<td>25</td>
<td>4-41</td>
<td>4 5</td>
<td>3 0 6</td>
<td>0/6 4/7 1/5</td>
<td>0/4</td>
<td>0/2 1/2</td>
</tr>
<tr>
<td>Third ventricle</td>
<td>3</td>
<td>2:1</td>
<td>19</td>
<td>14-33</td>
<td>1 2</td>
<td>1 1 1</td>
<td>0/2 1/2 0/1</td>
<td>0/2</td>
<td>0/1 0/1</td>
</tr>
<tr>
<td>Hypothalamus</td>
<td>1</td>
<td>1:0</td>
<td>30</td>
<td>-</td>
<td>1 0</td>
<td>0 0 1</td>
<td>0/1 0/1 1/1</td>
<td>-</td>
<td>0/1 0/1</td>
</tr>
<tr>
<td>Cerebellum</td>
<td>1</td>
<td>1:0</td>
<td>25</td>
<td>-</td>
<td>0 1</td>
<td>0 0 1</td>
<td>- - -</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>Pineal region</td>
<td>1</td>
<td>1:0</td>
<td>65</td>
<td>-</td>
<td>1 0</td>
<td>0 1 0</td>
<td>- 0/1 0/1</td>
<td>0/1</td>
<td>0/1 0/1</td>
</tr>
<tr>
<td>Total</td>
<td>47</td>
<td>25:22</td>
<td>14</td>
<td>2-65</td>
<td>34 13</td>
<td>10 14 23</td>
<td>0/30 15/32 14/33</td>
<td>1/18</td>
<td>0/15 4/15</td>
</tr>
</tbody>
</table>

Footnotes

WHO grade refers to histology at time of initial biopsy or resection.
p53+ refers to strong nuclear p53 staining in the majority of tumor cells.
EGFR amp refers to high level amplification of EGFR with >10 copies per cell.